

OXIDATION OF CORTICOSTEROIDS BY FLAVINS

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Abstract: The new reaction is reported between corticosteroids and flavins leading to the steroid-21-oic acids.

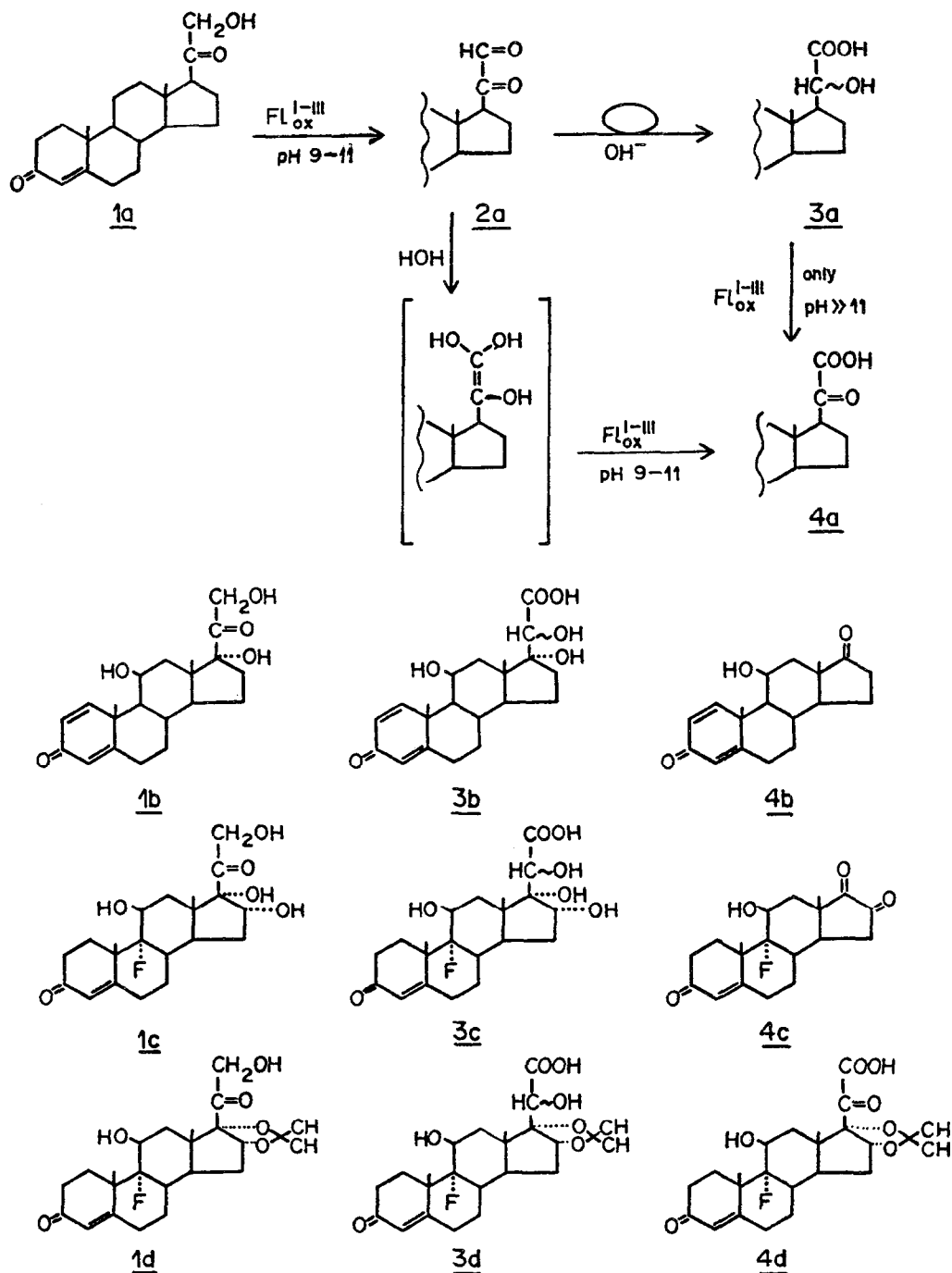
Until now, reports concerning synthesis of steroid 20-hydroxy-21-oic acids from the corticosteroids are still limited<sup>1-3</sup>. The possibility that corticosteroids are oxidized in vivo or in vitro to 21-oic acids has never been seriously considered. Only Monder et al. have shown that carboxylic acids are quantitatively important products of corticosteroid metabolism in man in vivo<sup>4</sup>.

In the present paper, we report results concerning synthesis in vitro of the respective steroid-21-oic acids from desoxycorticosterone 1a, prednisolone 1b, 9-fluoro-16 $\alpha$ -hydroxyhydrocortisone 1c, acetone of 9-fluoro-16 $\alpha$ -hydroxyhydrocortisone 1d in redox reaction with lumiflavin Fl<sub>Ox</sub><sup>I</sup>, 3-methylumiflavin Fl<sub>Ox</sub><sup>II</sup> and 8-carboxylumiflavin Fl<sub>Ox</sub><sup>III</sup>.

Synthesis of steroid acids: aqueous or aqueous/ethanol solutions of respective flavins (1mmol) and respective corticosteroids (0.5mmol) in modified Thunberg cuvette were deoxygenated by bubbling N<sub>2</sub> and reaction was achieved using 150W lamp. After irradiation stopped (0.5h) the solution were neutralised, then products isolated using column chromatography (silica gel, ethyl acetate - ethanol 3:1).

Under the conditions applied by us, in the oxidation of corticosteroids 1a-d by flavins Fl<sub>Ox</sub><sup>I-III</sup>, we obtained high yields of the respective steroid hydroxyacids 3a-d. We found that the oxidation proceeded by way of dehydrogenation of the  $\alpha$ -ketol group of the steroid, yielding glyoxal derivatives as intermediate products 2. In alkaline medium, as required for reaction with flavins (pH 9-11), the glyoxal 2 underwent predominantly an intramolecular Cannizzaro rearrangement, leading to the respective steroid hydroxyacids 3a-d.

In aqueous medium, we found that the glyoxalic derivatives 2 undergo partial hydration to give intermediate products containing en-diol forms. The en-diol system favours the re-dehydrogenation of these compounds in the presence of flavins leading to the  $\alpha$ -oxoacid 4a (from 1a), the 17-oxosteroid 4b (from 1b), the  $\alpha$ -dioxosteroid 4c (from 1c) and the  $\alpha$ -oxoacid 4d (from 1d).



In strongly alkaline medium ( $\text{pH} \gg 11$ ), the isolated steroid hydroxyacids  $3a-d$  partially underwent further oxygenation in presence of flavines to the derivatives  $4a-d$ , respectively. This reactions did not take place under standard conditions ( $\text{pH } 9-11$ ).

In the previous paper<sup>2</sup>, we pointed out the possibility of compounds of the type 4b and 4c arising by decomposition of hypothetical steroid hydroxy-oxo-acids, by analogy to formation of the stable  $\alpha$ -oxoacid 4a and acetonide of  $\alpha$ -oxoacid 4d. Acetonide blocking of the hydroxy groups at C-16 and C-17 (steroid 1d) lowers the lability of the side chain in corticosteroids. As a result, the side chain at C-17 is not cleaved in the oxidation of steroid 1d. This result is analogous to the result obtained from the oxidation of 1a which contains no hydroxyl group at C-16 or C-17, and is in contrast to the results of the oxidation of steroids 1b and 1c which contain hydroxy groups at C-17 and C-16 + C-17, respectively.

In table we give the yields of the oxidation products 3a-d and 4a-d.

Oxidizing agent	Potential <sup>5,6</sup> E <sub>m7</sub> (mV)	Yields (%) of oxidation products							
		<u>3a</u>	<u>3b</u>	<u>3c</u>	<u>3d</u>	<u>4a</u>	<u>4b</u>	<u>4c</u>	<u>4d</u>
Fl <sub>Ox</sub> <sup>I</sup>	- 0.207	41	37	40	32	16	8	11	15
Fl <sub>Ox</sub> <sup>II</sup>	- 0.223	46	43	38	34	11	15	20	14
Fl <sub>Ox</sub> <sup>III</sup>	- 0.457	67	56	52	46	9	12	10	19

The data in table show that in case when Fl<sub>Ox</sub><sup>III</sup> was applied as oxidizing agent, yields of steroid acids 3a-d are better than results obtained with Fl<sub>Ox</sub><sup>I</sup> and Fl<sub>Ox</sub><sup>II</sup>. This fact may be rationalized by the tendency of the reagents having a more negative potential to react at a higher rate<sup>7</sup>. When Fl<sub>Ox</sub><sup>III</sup> is used as the oxidizing agent, rapid dehydrogenation of the hydroxymethyl ketone take place with formation of the intermediate 20,21-dioxo derivative 2. Hence, the rate of this reaction stage essentially controls the yield of the steroid hydroxyacid 3 which is formed from the 20,21-dioxo derivative 2 by an intramolecular Cannizzaro reaction. Detailed results of the studies on the mechanism of this redox reaction will published later.

Physical data for oxidation products:

- their structure was confirmed by <sup>1</sup>H-NMR(DMSO-d<sub>6</sub>, TMS,  $\delta$  ppm) and IR (cm<sup>-1</sup>),
- all compounds gave satisfactory elemental analysis,
- melting points are uncorrected.

3a (20-hydroxy-3-oxo-pregn-4-en-21-oic acid) : m.p. 160-163°C,  
molecular formula C<sub>21</sub>H<sub>30</sub>O<sub>4</sub> (346.5); IR: 3395, 1725, 1660, 1615, 1265;  
NMR: 0.7(s, 18-CH<sub>3</sub>), 1.20(s, 19-CH<sub>3</sub>), 3.4-3.7(m, OH-acid), 5.8(s, 1H, H-C-4).

4a (3,20-dioxopregn-4-en-21-oic acid) : m.p. 157-161°C,  
molecular formula C<sub>21</sub>H<sub>28</sub>O<sub>4</sub> (344.4); IR: 3050, 1730, 1715, 1660, 1250;  
NMR: 0.75(s, 18-CH<sub>3</sub>), 1.15(s, 19-CH<sub>3</sub>), 4.4(s, OH-acid), 5.75(s, 1H, H-C-4).

- 3b (11 $\beta$ ,17 $\alpha$ ,20-trihydroxy-3-oxopregn-1,4-dien-21-oic acid) : m.p. 235-238°C, molecular formula C<sub>21</sub>H<sub>28</sub>O<sub>4</sub> (376.4); IR: 3390, 1730, 1660, 1600, 1250; NMR: 1.0(s,18-CH<sub>3</sub>), 1.4(s,19-CH<sub>3</sub>), 3.8-4.1(m,OH-acid), 4.5(m,1H,H-C-11), 5.9(s), 6.1(d), 7.3(d,H-C-4,H-C-1).
- 4b (11 $\beta$ -hydroxy-3,17-dioxoandrost-1,4-diene) : m.p. 157-158°C, molecular formula C<sub>19</sub>H<sub>24</sub>O<sub>3</sub> (300.4); IR: 3400, 1735, 1655, 1610; NMR: 0.95(s,18-CH<sub>3</sub>), 1.5(s,19-CH<sub>3</sub>), 4.6(m,1H,H-C-11), 5.9(s), 6.1(d), 7.3(d,H-C-4,H-C-1).
- 3c (9-fluoro-11 $\beta$ ,16 $\alpha$ ,17 $\alpha$ ,20-tetrahydroxy-3-oxopregn-4-en-21-oic acid) : m.p. 261-263°C, molecular formula C<sub>21</sub>H<sub>29</sub>FO<sub>7</sub> (412.5); IR: 3320, 2610, 1735, 1660, 1615; NMR: 1.1(s,18-CH<sub>3</sub>), 1.4(s,19-CH<sub>3</sub>), 3.9-4.2(m,OH-acid), 4.8(m,H-C-11), 5.75(s,H-C-4).
- 4c (9-fluoro-11 $\beta$ -hydroxy-3,16,17-trioxoandrost-4-ene) : m.p. 225-227°C, molecular formula C<sub>19</sub>H<sub>23</sub>FO<sub>4</sub> (334.4); IR: 3400, 1710-1725, 1660, 1615; NMR: 0.95(s,18-CH<sub>3</sub>), 1.4(s,19-CH<sub>3</sub>), 4.7(m,1H,H-C-11), 5.75(s,H-C-4).
- 3d (O,O-isopropylidene-9-fluoro-11,20-dihydroxy-3-oxopregn-4-en-21-oic acid) : m.p. 267-270°C, molecular formula C<sub>24</sub>H<sub>33</sub>FO<sub>7</sub> (452.5); IR: 3380, 2690, 1735, 1660, 1615sh, 1245, 1055; NMR: 1.1(s,18-CH<sub>3</sub>), 1.5-1.7(m,3CH<sub>3</sub>), 4.05-4.15(m,OH-acid), 4.7(m,1H,H-C-11), 5.75(s,H-C-4).
- 4d (O,O-isopropylidene-9-fluoro-11-hydroxy-3,20-dioxopregn-4-en-21-oic acid) : m.p. 252-254°C, molecular formula C<sub>24</sub>H<sub>31</sub>FO<sub>7</sub> (450.5); IR: 3500, 1735sh, 1725, 1660, 1620sh, 1240, 1070; NMR: 1.1(s,18-CH<sub>3</sub>), 1.6-1.8(m,3CH<sub>3</sub>), 4.9(m,1H,H-C-11), 5.8(s,H-C-4).

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#### References:

1. J.Jasiczak, M.A.Smoczkiewicz, Zesz.Nauk. AE-Poznan 1973, 53, 125; C.A. 81, 136360 (1974).
2. J.Jasiczak, M.A.Smoczkiewicz, Zesz.Nauk. AE-Poznan 1978, 80, 76; C.A. 93, 186649 (1980).
3. M.A.Smoczkiewicz, J.Jasiczak, Synthesis 1980, 739.
4. C.Monder, K.O.Martin, Biochemistry 1976, 15, 576.
5. W.Mansfield Clark, "Oxidation-Reduction Potentials of Organic Systems", Waverly Press, Inc. Baltimore 1960, pp. 441-447.
6. S.Kasai, Y.Kubo, S.Yamanaka, J.Nutr.Sci.Vitaminol., 24, 339, 1978.
7. F.P.Altman, "An Introduction to Quantitative Enzyme Cytochemistry", Koch-Light Lab. Ltd., Colnbrook, England, 1972.

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